

Pollen Grain Variation among Some Trees of Fabaceae in New Damietta, Egypt

Amina. Z. Abo-Elnaga¹, Ahmed Ali*¹ and Sami Rabei¹

¹Botany and Microbiology Department, Faculty of Science, Damietta University, New Damietta 34517, Egypt.

Received: 24 June 2022 /Accepted: 14 August 2022

* Corresponding author's E-mail: ahmed.salama@du.edu.eg

Abstract

The current study aims at investigating the morphology of pollen grain for eleven woody species of Fabaceae cultivated in New Damietta, Egypt. The pollen grains were prepared according to Erdtman's acetolysis method, described and photographed using light microscopy (LM). The results showed variations in their qualitative and quantitative features. Differences in pollen shape (subprolate, prolate, prolate-spheroidal, oblate-spheroidal), size (medium to large), pollen arrangement (monads or polyads), amb (circular, irregular, elliptic or triangular), polarity (isopolar, heteropolar), type of apertures (porate, colporate, inaperturate) and exine ornamentation patterns (psilate, verrucate, granulate, reticulate and psilate-perforate) are characterized. Electrophoretic (SDS-PAGE) examination of total pollen protein content of four species (*Acacia ehrenbergiana*, *Cassia glauca*, *Delonix regia*, *Leucaena leucocephala*) revealed distinct protein bands. The results of this study is of great importance in understanding the pollen types of some allergy producing species.

Keywords: Egypt, Fabaceae, New Damietta, Pollen grains morphology.

Introduction

The Fabaceae (or Leguminosae) family is among the most varied families, with members found in almost all ecological systems on the earth (Bahadur et al., 2022). The third largest angiosperm family, with species that are morphologically, physiologically, and environmentally diverse (APG III 2009; Schwarz et al., 2015; Werner et al., 2015). Cercidoideae, Detarioideae, Duparquetioideae,

Dialioideae, Caesalpinioideae, and Papilionoideae are the six subfamilies of the Fabaceae. The biggest subfamily, Papilionoideae, has 30 tribes, 455 genera, and over 12,000 species (LPWG, 2017). It is the most widespread, and its members are adaptable to a wide range of habitats.

Fabaceae is eurypalynous family (Luz et al., 2013), with vastly diverse pollen characters (Pavlova & Manova, 2000; Taia, 2004; Lashin 2006; Liao et al., 2022). Among the numerous studies that explain the pollen

staining, the gels were dried. The identity of protein bands was determined by their molecular weights based on the loaded protein ladder (PageRuler™ Prestained Protein Ladder, Thermo Fisher Scientific, Cat. No. 26616).

Table (1): List of selected Fabaceae species with their localities.

Taxon	Geography coordinates	Life form	Life span
<i>Acacia ehrenbergiana</i>	31°26'□22.90"N 31°40'□56.05"E	Evergreen small tree	Perennial
<i>Acacia saligna</i>	31°26'□20.35"N 31°40'□57.87"E	Evergreen shrub or small tree	Perennial
<i>Cassia alata</i>	31°26'□23.71"N 31°40'□59.68"E	Evergreen tree	Perennial
<i>Calliandra emarginata</i>	31°26'□21.89"N 31°40'□59.68"E	Evergreen small tree	Perennial
<i>Cassia fistula</i>	31°26'□21.85"N 31°41'□00.17"E	Deciduous tree	Perennial
<i>Cassia glauca</i>	31°26'□29.55"N 31°41'□00.36"E	Deciduous tree	Perennial
<i>Cassia nodosa</i>	31°26'□21.89"N 31°40'□59.68"E	Deciduous tree	Perennial
<i>Cassia tora</i>	31°26'□28.80"N 31°41'□01.14"E	Deciduous shrub or small tree	Perennial
<i>Delonix regia</i>	31°26'□21"N 31°40'□59"E	Deciduous tree	Perennial
<i>Leucaena leucocephala</i>	31°26'□23.53"N 31°40'□59.61"E	Evergreen shrub or small tree	Perennial
<i>Tipuana tipu</i>	31°26'□06.60"N 31°41'□04.13"E	Deciduous tree	Perennial

Principal component analysis

Principal component analysis (PCA) ordination was performed using the program PAST version (4.04). The characteristics used for PCA included: Pollen unit, polarity, pollen shape, aperture type, exine sculpture, polar axis, equatorial diameter and pollen size.

Results

The palynological data of the examined species are listed alphabetically by genus and species, abbreviations and measurements are placed within brackets, and all data are reported in table (3).

Pollen grains morphology

***Acacia ehrenbergiana* Hayne.** Pollen grains isopolar, subprolate in equatorial view, large in size with mean dimensions of (44.46 - 55.4) μm. The grains are inaperturate, exine psilate, exine thickness 1.6 μm, P/E ratio 1.25, outline in polar view rounded.

***Acacia saligna* Labill.) H.L. Wendl.** Pollen grains isopolar, subprolate in equatorial view, medium in size with mean dimensions of (36.8 - 48.5) μm. The grains are inaperturate, exine psilate, exine thickness 1.46 μm, P/E ratio 1.32, outline in polar view circular to semi-rounded.

***Cassia alata* L.** Pollen grains isopolar, prolate-spheroidal in equatorial view, medium in size with mean dimensions of (29 - 29.9) μm. The grains are tricolporate, exine psilate, exine thickness 1.59 μm, P/E ratio 1.03, outline in polar view circular.

***Calliandra emarginata* Benth.** Pollen grains heteropolar, prolate in equatorial view, very large in size with mean dimensions of (68 - 114) μm. Polyads calymmate, with one end rounded and the opposite end tapered, composed of 8 pollen grains, being one central pollen grain surrounded by seven peripheral heteromorphic pollen grains, one of the peripheral has a conical and tapered shape. The grains are tetra- to penta-porate, exine verrucate, exine thickness 2.2 μm, P/E ratio 1.676, outline in polar view irregular- elliptic.

***Cassia fistula* L.** Pollen grains isopolar, oblate-spheroidal in equatorial view, medium in size with mean dimensions of (33.8 - 35.9) μm. The grains are tricolporate, exine granulate, exine thickness 2.8 μm, P/E ratio 0.94, outline in polar view circular to subtriangular.

***Cassia glauca* L.** Pollen grains isopolar, prolate-spheroidal in equatorial view, medium in size with mean dimensions of (37 - 37.75) μm. The grains are tricolporate, exine psilate, exine thickness 2.03 μm, P/E ratio 1.02, outline in polar view circular- elliptic.

***Cassia nodosa* L.** Pollen grains isopolar, prolate-spheroidal in equatorial view, medium in size with mean dimensions of (27.7 - 30.5) μm. The grains are tricolporate, exine psilate, exine thickness 1.84 μm, P/E ratio 1.1, outline in polar view triangular.

***Cassia tora* (L.) Roxb.** Pollen grains isopolar, prolate-spheroidal in equatorial view, medium in size with mean dimensions of (25.59 - 27.5) μm. The grains are triporate, exine psilate, exine thickness 2 μm, P/E ratio 1.072, outline in polar view circular- elliptic.

***Delonix regia* (Boj. ex Hook) Raf.** Pollen grains isopolar, subprolate in equatorial view, medium in size with mean dimensions of (33.3 - 36.3) μm. The grains are tricolporate, exine reticulate, exine thickness 4.56 μm, P/E ratio 1.09, outline in polar view circular.

Leucaena leucocephala (Lam.) de wit. Pollen grains isopolar, prolate-spheroidal in equatorial view, medium in size with mean dimensions of (31.31 - 34.39) μm . The grains are tricolporate, exine psilate-perforate, exine thickness 2.97 μm , P/E ratio 1.098, outline in polar view circular.

Tipuana tipu (Benth.) Kuntze. Pollen grains isopolar, prolate-spheroidal in equatorial view, medium in size with mean dimensions of (25.2 - 27) μm . The grains are tricolporate, exine psilate, exine thickness 1.55 μm , P/E ratio 1.07, outline in polar view triangular.

Pollen protein analysis

Electrophoretic (SDS-PAGE) analysis of total pollen protein content revealed several different protein bands. Two bands with molecular weights of 45 and 72 kDa in *Acacia ehrenbergiana*, four bands with molecular weights of 32, 55, 64, 95 kDa in *Cassia glauca*, two bands with molecular weights of 20 and 55-250 kDa in *Delonix regia*, two intensive bands with molecular weights range between 28-34 kDa and 40-95 kDa in *Leucaena leucocephala*.

Table (2): Representation of protein bands and their equivalent molecular weight (KDa) of some selected species of Fabaceae.

Scientific name	MWT (KDa)
<i>A. ehrenbergiana</i>	45
	72
<i>C. glauca</i>	32
	55
	64
	95
<i>D. regia</i>	20
	55-250
<i>L. leucocephala</i>	28-34
	40-95

PLATE (A)

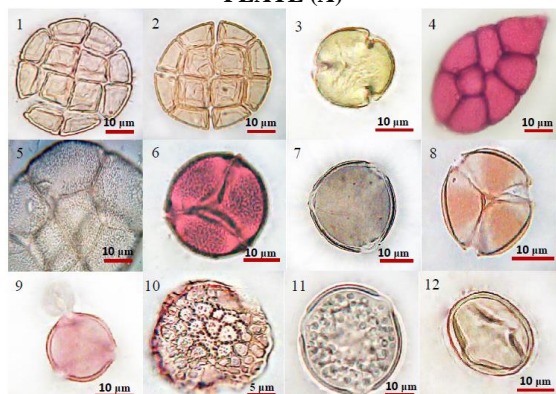


PLATE (A): Light micrographs of pollen grains of *A. ehrenbergiana* (1), *A. saligna* (2), *C. alata* (3), *C. emarginata* (4,5), *C. fistula* (6), *C. glauca* (7), *C. nodosa* (8), *C. tora* (9), *D. regia* (10), *L. leucocephala* (11), *T. tipu* (12).

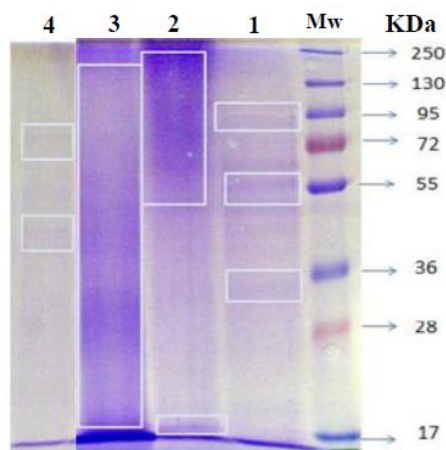


Figure (2): SDS-PAGE protein profiling of pollen (Molecular weight of bands assigned based on the similarity with molecular weight marker). (1- *C. glauca*, 2- *D. regia*, 3- *L. leucocephala*, 4- *A. ehrenbergiana*).

The palyno-morphological characteristics of 11 species belonging to 6 genera of the Fabaceae family are shown in Table 1. Photographs of the examined species were collected in (Plate B). Variations in all of the examined species have been reported using both quantitative and qualitative features. Polar and equatorial diameter (Figure 3), aperture types (Figure 4), P/E ratio (Figure 5), and exine thickness (Figure 6).

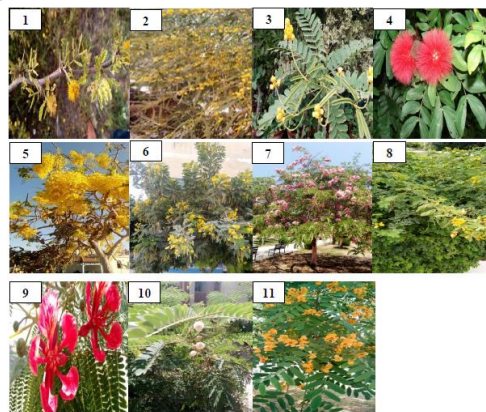


PLATE (B): Plant species of family Fabaceae in New Damietta, (1) *A. ehrenbergiana*, (2) *A. saligna*, (3) *C. alata*, (4) *C. emarginata*, (5) *C. fistula*, (6) *C. glauca*, (7) *C. nodosa*, (8) *C. tora*, (9) *D. regia*, (10) *L. leucocephala*, (11) *T. tipu*.

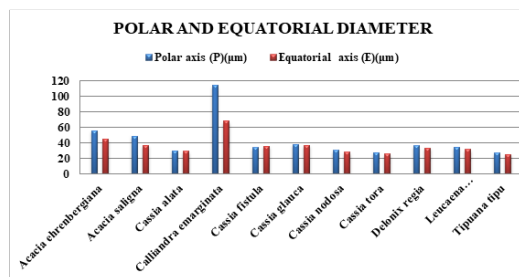


Figure (3): Polar and equatorial diameter variations of some selected Fabaceae species.

VARIATION IN APERTURE TYPE

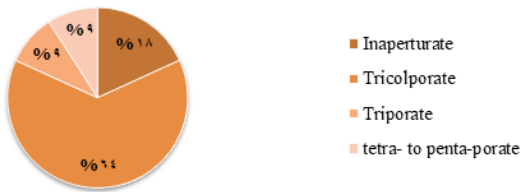


Figure (4): Aperture variations of selected Fabaceae species

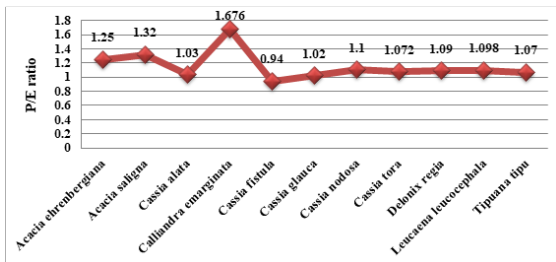


Figure (5): P/E index of some selected Fabaceae species

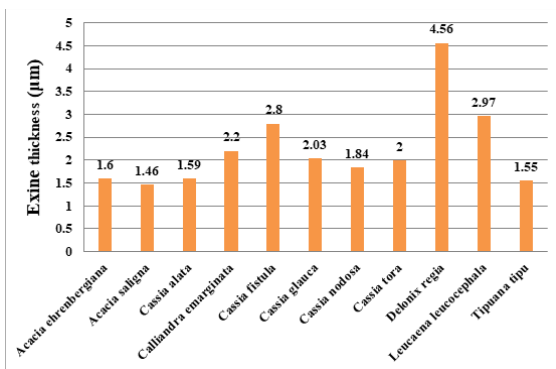


Figure (6): Variation in exine thickness of some selected Fabaceae species.

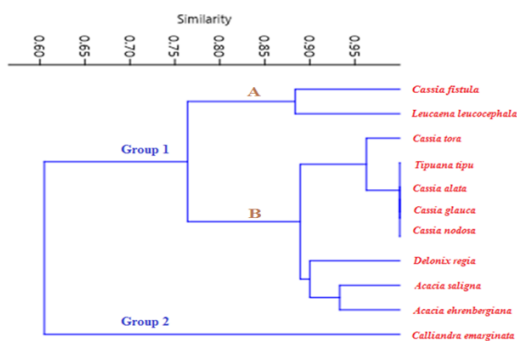


Figure (7): Dendrogram based on the palynological characters of some selected Fabaceae species.

Table (3): PCA of some selected species of Fabaceae

PC	Eigenvalue	% variance
1	11.3712	84.516
2	1.71377	12.738
3	0.151943	1.1293
4	0.0955564	0.71022
5	0.0887068	0.65931

6	0.0232064	0.17248
7	0.0101139	0.075171
8	3.27776E-34	2.4362E-33

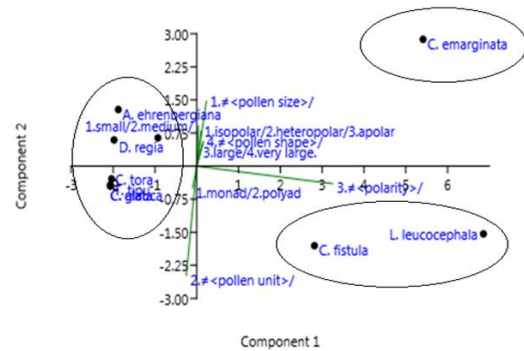


Figure (8): PCA scatter biplot of selected species of Fabaceae

Discussion

Light microscopic examinations of 11 species from the Fabaceae family in New Damietta province provided useful information on pollen morphological features. Pollen shape, size, polarity, polar and equatorial outlines, exine thickness, number of apertures, and exine ornamentation are the most important morphological aspects of the species studied. The pollen grains from Fabaceae family presented in this study show variation in terms of pollen unit, size, amb, shape, polarity, exine thickness, and ornamentation. These variations were also identified in the descriptions of certain species in results of NIEZGODA et al., (1983), Maghni (2020), Deshmukh et al., (2014), Khan et al., (2021) and Ullah et al., (2022).

The results showed that the shape of pollen grain is subprolate, rounded or circular for *A. ehrenbergiana*, *A. saligna*. This result confirms that stated by Al-Watban et al., (2013) and Rajurkar et al., (2013) with 16 monads which was similar to that revealed by Caccavari and Dome (2000). *Delonix regia* has subprolate, tricolporate and reticulate exine which was in agreement with the results of Orijemie, (2017) and Ullah et al., (2022) but it conflicted with that was described by Antonio-Domingues et al., (2018) who revealed oblate-spheroidal shape. Prolate-spheroidal shape recorded in *Cassia alata*, *Cassia glauca*, *Cassia nodosa*, *Cassia tora*, *Leucaena leucocephala* and

Table (4): The pollen morphological characters of the examined taxa (values in parentheses represent average lengths)

Taxa	Polar axis (P)(µm)	Equatorial axis (E)(µm)	P/E ratio	Pollen Size	Pollen Unit	Shape in Equatorial view	Shape in Polar view	Polarity	Aperture condition	Exine thickness (µm)	Exine sculpture Pattern
<i>Acacia ehrenbergiana</i>	50.7-58.5 (55.4±3.3)	27.3-54.6 (44.5±10.5)	(1.25) 1.07-1.27	Large	Polyad	Subprolate	Rounded	Isopolar	Inaperturate	1.6	Psilate
<i>Acacia saligna</i>	44-52 (48.5±2.9)	28-48 (36.8±5.8)	(1.32) 1-1.85	Medium	Polyad	Subprolate	Circular to Semi-rounded	Isopolar	Inaperturate	1.46	Psilate
<i>Cassia alata</i>	27.9-32.3 (29.9±1.7)	27.9-29.4 (29± 0.69)	(1.03) 1-1.09	Medium	Monad	Prolate Spheroidal	Circular	Isopolar	Tricolporate	1.59	Psilate
<i>Calliantha emarginata</i>	100-130 (114±9.8)	60-70 (68±4.1)	(1.676) 1.43-1.86	Very large	Polyad	Prolate	Irregular-Elliptic	Heteropolar	Tetra- to pentaporate	2.2	Verrucate
<i>Cassia fistula</i>	27.9-37 (33.8±2.3)	31.2-39 (35.9± 2.7)	(0.94) 0.8-1.13	Medium	Monad	Oblate Spheroidal	Circular to subtriangular	Isopolar	Tricolporate	2.8	Granulate
<i>Cassia glauca</i>	35.3-41.2 (37.75 ± 2)	35.3-38.2 (37±1.4)	(1.02) 1-1.08	Medium	Monad	Prolate Spheroidal	Circular-Elliptic	Isopolar	Tricolporate	2.03	Psilate
<i>Cassia nodosa</i>	23.4-35.1 (30.5±2.9)	23.4-35.1 (27.7±2.8)	(1.1) 0.86-1.3	Medium	Monad	Prolate Spheroidal	Triangular	Isopolar	Tricolporate	1.84	Psilate
<i>Cassia tora</i>	26.2-27.8 (27.5±0.7)	23.2-27.72 (25.59±1.7)	(1.072) 1-1.2	Medium	Monad	Prolate Spheroidal	Circular-Elliptic	Isopolar	Triporate	2	Psilate
<i>Delonix regia</i>	32.5-42.5 (36.3±4.5)	30-42.5 (33.3±4.4)	(1.09) 1-1.3	Large	Monad	Subprolate	Circular	Isopolar	Tricolporate	4.56	Reticulate
<i>Leucaena leucocephala</i>	29.4-38.2 (34.4±3.03)	26.46-38.2 (31.31±3.7)	(1.098) 1-1.19	Medium	Monad	Prolate Spheroidal	Circular	Isopolar	Tricolporate	2.97	Psilate-Perforate
<i>Tipuana tipu</i>	22.05-30.8 (27± 3.8)	19.2-32.34 (25.2±4.7)	(1.07) 0.88-1.4	Medium	Monad	Prolate Spheroidal	Triangular	Isopolar	Tricolporate	1.55	Psilate

Tipuana tipu. Prolate shape for *C. emarginata*, oblate-spheroidal for *C. fistula* which disagrees with that stated by Ullah et al., (2022). Also, the psilate exine was found in seven species, *A. ehrenbergiana*, *A. saligna*, *C. alata*, *C. glauca*, *C. nodosa*, *C. tora* and *T. tipu*. Verrucate, granulate, reticulate and psilate-perforate patterns are represented by one species only as follow *C. emarginata*, *C. fistula*, *D. regia* and *L. leucocephala* respectively. In comparison with previous studies: *L. leucocephala* has tricolpate and subpsilate to scabrate exine sculpturing (Aftab & Perveen, 2006 ; Bahadur et al., 2022), this was not supported by our findings. The tricolporate pollen grains are found in seven species, *C. alata*, *C. glauca*, *C. nodosa*, *C. fistula*, *D. regia*, *L. leucocephala* and *T. tipu*. Porate type is found in two species, *C. emarginata* and *C. tora*. Inaperturate pollen grains are found in *A. ehrenbergiana*, *A. saligna* which corresponded with the results of Maghni, (2020). But it disagrees with that reported by Al-Watban et al., (2013) since *Acacia* species have colpate pollen grains. Qualitative and quantitative characteristics indicated that some of the species were compatible with previous studies and exhibit similarities in traits, demonstrating the genus' consistency.

The cluster analysis (Dendrogram) at 60% similarity showed two distinct groups of species (Groups 1 and 2) in Figure (7); Group (1) includes ten species divided into two groups (A & B) at 75% similarity, group (A) shows \approx 88% similarity and consists of two species: *C. fistula* and *L. leucocephala*; group (B) contains the reminder of species and shows \approx 88% similarity was further divided into two groups. The first group contained five species at 95% similarity: *C. tora*, *T. tipu*, *C. alata*, *C. glauca* and *C. nodosa*. The second group included three species: *A. ehrenbergiana*, *A. saligna*, *D. regia*. Group 2 comprises of one species: *C. emarginata*. This grouping was also reflected in the PCA. The pollen morphology of the Fabaceae taxa studied demonstrated high consistency, with just minor variations across species.

The results identified an allergenic protein band with a molecular mass of 45 KDa in *A. ehrenbergiana*, which was consistent with a previous result of (Shamsbiranvand, et al., 2014). This band was also detected in *L. leucocephala*, suggesting that the 45 kDa

protein identified in this study could be a candidate for being one of the proteins responsible for the allergy in this species. There were six major allergenic proteins in *Phoenix dactylifera* with molecular mass ranging from 12 to 94 kDa bands (Kwaasi et al., 1993; Asturias et al., 2005). Among these proteins, there was a 55 kDa band discovered in *Cassia glauca*, *Delonix regia*, *Leucaena leucocephala*. The 20-kDa band was also identified in *D. regia*. Protein bands of 28-30 kDa were also identified in *L. leucocephala*. According to the findings *A. ehrenbergiana*, *C. glauca*, *D. regia*, and *L. leucocephala* contain molecular weight related proteins which may cause allergies in these plants, but further research utilizing immunoblotting analysis could be required to confirm these results.

Taxonomic key based on pollen micro-morphology

- 1- Pollen in polyads 2
- Pollen in Monads..... 4
- 2- Polyads with one tapered end, composed of 8 grains *Calliandra emarginata*
- Polyads with uniform ends, composed of 16 grains 3
- 3- Polyads medium sized (44-52 μ m x 38–48 μ m), *Acacia saligna*
- Polyads large in size (50.7-58.5 μ m x 27.3-54.6 μ m), *Acacia ehrenbergiana*
- 4- Pollen grains oblate spheroidal or subprolate 5
- Pollen grains prolate spheroidal 6
- 5- Pollen grains oblate spheroidal, exine granulate *Cassia fistula*
- Pollen grains subprolate, exine reticulate
- Delonix regia*
- 6- Pollen grain triporate *Cassia tora*
- Pollen grain tricolporate 7
- 7- Exine sculpture is psilate 8
- Exine sculpture is psilate- perforate
- *Leucaena leucocephala*
- 8- Exine thickness more than 1.8 μ m *Cassia glauca & Cassia nodosa*
- Exine thickness less than 1.8 μ m
- *Cassia alata & Tipuana tipu*

References

Aftab, R.; Perveen, A. (2006). A palynological study

- of some cultivated trees from Karachi. Pak. J. Bot., 38: 15–28.
- Al-Watban, A. A., Al-Mogren, E., Doaigey, A. R., & El Zaidy, M. (2013). Pollen morphology of seven wild species of *Acacia* in Saudi Arabia. *African Journal of Plant Science*, 602-607.
- Antonio-Domingues H, Corrêa AMS, Queiroz RT, Bitar NAB. (2018). Pollen morphology of some Fabaceae species from Patos de Minas, Minas Gerais State, Brazil. *Hoehnea*, 45:103–114.
- APG III. (2009). An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants. *Bot J Linn Soc.*, 161: 105-202.
- Asher, M.I., Montefort, S., Bjorksten, B., Lai, C.K., Strachan, D.P., Weiland, S.K., et al.(2006). Worldwide Time Trends in the Prevalence of Symptoms of Asthma, Allergic Rhinoconjunctivitis, and Eczema in Childhood: ISAAC Phases One and Three repeat Multicountry Cross-Sectional Surveys. *Lancet*, 368, 733-743.
- Asturias, J A, I Ibarrola, J Fernández, M C Arilla, R González-Rioja, and A Martínez. (2005). Pho d 2, a major allergen from date palm pollen, is a profilin: cloning, sequencing, and immunoglobulin E cross-reactivity with other profilins. *Clin Exp Allergy*, 35:374–381.
- Bahadur, S., Taj, S., Long, W., Hanif, U. (2022). Pollen Morphological Peculiarities of Selected Mimosoideae Taxa of Hainan Island and Their Taxonomic Relevance. *Agronomy*, 1-13.
- Banks H, Lewis G. (2009). Pollen morphology of the *Dimorphandra* group (Leguminosae, Caesalpinioideae). *Grana*, 48:19–26.
- Barth OM, Côte-Real S, Macieira EG. (1976). Morfologia do polén anemófilo e alergizante no Brasil: II. Memórias Do Instituto Oswaldo Cruz. 74(3–4):191–201.
- Bradford, M.M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72: 248–254.
- D'Amato, G., Cecchi, L., Bonini, S., Nunes, C., Annesi-Maesano, I., Behrendt, H., et al. (2007). Allergenic pollen and pollen allergy in Europe. *Allergy*, 62: 976–990.
- Deshmukh A S, Barge H S and Gaikwad K D. (2014). PALYNO MORPHOMETRIC STUDIES IN SOME CASSIA L. SPECIES FROM MAHARASHTRA. *Indian Journal of Plant Sciences*, 71-78.
- Eduardo Lopes Soares, Lorryne Albernaz Domingues Camilo Landi, Cintia Neves De Souza, Eduardo Custódio Gasparino. (2022). Polyads types of the mimosoid clade (Caesalpinioideae, Fabaceae): size and pollen numbers variations. *Grana*, 61:45-66.
- Erdtman, G. (1952). *Pollen Morphology and Plant Taxonomy (An Introduction to Palynology, Angiosperms)* Chronica Botanica Co, Waltham, Mass/ Almqvist and Wicksell, Stockholm, 539 pp.
- Erdtman, G. (1960). The acetolysis method: a revised description. *Svensk Bot Tidskr*, 54:561–564.
- Faegri K, Iversen J. (1989). *Textbook of pollen analysis*. 4th edition: Faegri K, Kaland PE, and Krzywinski K. John Wiley, Chichester.
- Graham A, Barker G, Silva MF. (1980). Unique Pollen Types in the Caesalpinioideae (Leguminosae). *Grana*, 19:79–84.
- Khan SU, Zafar M, Ullah R, et al. (2021). Pollen diversity and its implications to the systematics of mimosaceous species by LM and SEM. *Microsc Res Tech.*, 84:42–55.
- Kwaasi, AAA, R S Parhar, P Tipirneni, H Harfi, and S T Al-Sedairy. (1993). Major allergens of date palm (*Phoenix dactylifera* L.) pollen. *Allergy*, 48: 511-518.
- Laemmli, E.K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227:680–685.
- Lashin, G.M. (2006). Comparative morphology of pollen grains of some taxa of tribe Trifolieae (Fabaceae: Papilionoideae) from Egypt. *Int J Bot.*, 2: 210-211.
- Liao, M., Ullah, F., Deng, H.-N., Zhang, J.-Y., Xu, B., & Gao, X.-F. (2022). Pollen morphology of the genus *Sophora* (Fabaceae) and its taxonomic implications. *Microscopy Research and Technique*, 85(5), 1723–1741.
- LPWG, (2017). A new subfamily classification of the Leguminosae based on a taxonomically comprehensive phylogeny. *Taxon*, 66: 44-77.
- Luz CFP, Maki ES, Horák-Terra I, Vidal-Torrado P, Mendonça Filho CV. (2013). Pollen grain morphology of Fabaceae in the Special Protection Area (SPA) Pau-de-Fruta, Diamantina, Minas Gerais, Brazil. *An Acad Bras Cienc*, 85:1329–1344.
- Maghni B, Ait Hammou M, Khedim R., Maatoug M, Hellal B. (2020). Palynological study of angiosperms of rostomid park of Tiaret in Algeria. *J. Fundam. Appl. Sci.*, 12(1), 1-11.
- Niezgoda Christine J, Feuer s Sylvia M and Nevling Lorin I. (1983). Pollen ultrastructure of the Tribe Ingeae (Mimosoideae: Leguminosae). *Amer. J. Bot.*, 70: 650-667.
- Orijemie, E. A. (2017). Pollen morphology of Tribe Caesalpinioideae (Leguminosae) ornamental species in Nigeria. *Nigerian Journal of Botany*, 31, 273-283.

- Pavlova, D.K., Manova, V.I. (2000). Pollen morphology of the genera *Onobrychis* and *Hedysarum* (Hedysareae, Fabaceae) in Bulgaria, *Ann. Bot. Fenn.*, 207-217.
- POWO. (2022). "Plants of the World Online. Facilitated by the Royal Botanic Gardens, Kew. Published on the Internet; <http://www.plantsoftheworldonline.org/Retrieve> d 20 October 2021."
- Punt W, Hoen PP, Blackmore S, Nilsson S, Le Thomas A. (2007). Glossary of pollen and spore terminology. *Rev Palaeobot-Palynol.*, 143(1-2):1-81.
- Schwarz, E.N., Ruhlman, T.A., Sabir, J.S., Hajrah, N.H., Alharbi, N.S., Al-Malki, A.L., Bailey, C.D., Jansen, R.K. (2015). Plastid genome sequences of legumes reveal parallel inversions and multiple losses of *rps16* in papilionoids. *J Syst Evol.*, 53: 458-468.
- Shamsbiranvand, Mohammad Hosein , Ali Khodadadi, Mohammad Ali Assarehzadegan, Seyed Hamid Borsi, and Akram Amini. (2014). Immunochemical Characterization of *Acacia* Pollen Allergens and Evaluation of Cross-Reactivity Pattern with the Common Allergenic Pollens. *Journal of Allergy*, 1-7.
- Taia, W.K. (2004). Palynological study within tribe *Trifolieae* (Leguminosae). *Pakistan J. Biol. Sci.*, 7: 1303-1315.
- Taia, W.K. (2020). Pollen Allergens of some Road Trees, Shrubs and Herbs in Alexandria, Egypt. 2 (1) *OAJBS.ID.000143*.
- Talebi, S. M., Azizi, N., Yadegari, P., & Matsyura, A. (2020). Analysis of pollen morphological characteristics in Iranian *Onobrychis* Miller (Fabaceae) taxa. *Brazilian Journal of Botany*, 609-632.
- Tütüncü Konyar, S. and Dane, F. (2012). Pollen Morphology of Exotic Trees and Shrubs of Edirne II. *Journal of Applied Biological Sciences*, 6 (2): 13-18.
- Ullah, F., Ahmad, M., Zafar, M., Parveen, B., Ashfaq, S., Bahadur, S., Safdar, Q., Safdar, L. B., Alam, F., & Luqman, M. (2022). Pollen morphology and its taxonomic potential in some selected taxa of *Caesalpiniaceae* observed under light microscopy and scanning electron microscopy. *Microscopy Research and Technique*, 85(4), 1410-1420.
- Werner, G.D., Cornwell, W.K., Cornelissen, J.H., Kiers, E.T. (2015). Evolutionary signals of symbiotic persistence in the legume-rhizobia mutualism. *P Natl Acad Sci.*, 112: 10262-10269.

الملخص العربي

عنوان البحث:

أمينة أبو النجا¹، أحمد علي*¹، سامي ربيع¹
¹ قسم النبات والميكروبيولوجي، كلية العلوم، جامعة دمياط، دمياط، مصر.

تهدف الدراسة الحالية إلى التحقيق في مورفولوجيا حبوب اللقاح لأحد عشر نوعاً خشبياً من الفصيلة القرنيّة المزروعة في مدينة دمياط الجديدة، تم اعداد شرائح حبوب اللقاح وذلك بعد صباغتها طبقاً لطريقة (Erdtman 1960) ودراسة الصفات الباليولوجية المختلفة وتصويرها باستخدام المجهر الضوئي، وأظهرت النتائج تبايناً في خصائص حبوب اللقاح النوعية والكمية ومن خلال هذه الدراسة أمكن وضع مفتاحاً يمكن بواسطته التعرف والتفرقة بين هذه النباتات التي تم دراستها. بالإضافة إلى دراسة بعض حبوب اللقاح المسببة للحساسية عن طريق فصل البروتين في هذه العينات باستخدام الفصل الكهربائي (SDS-PAGE analysis) لإجمالي محتوى بروتين حبوب اللقاح في أربعة أنواع، وكانت النتيجة هي ظهور **Protein bands** ذات أوزان جزيئية تتراوح بين 20-250 كيلودالتون في الأربعة الأنواع.